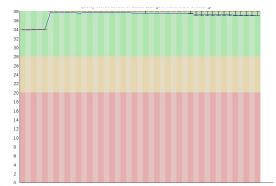
# FastQC

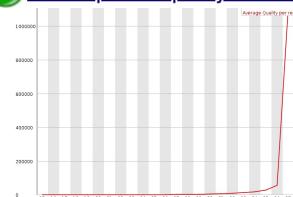
## FastQC

#### Quality

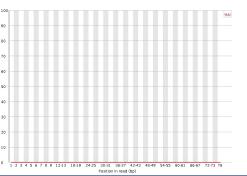
Per base sequence quality



Per sequence quality scores

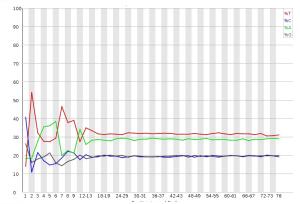


Per base N content

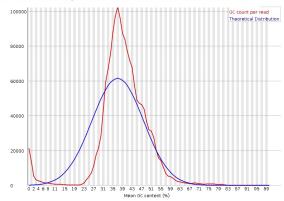


#### **Characteristics**

Per base sequence content



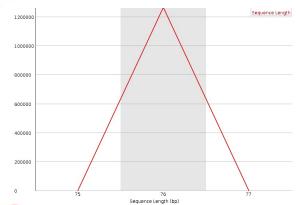
Per sequence GC content



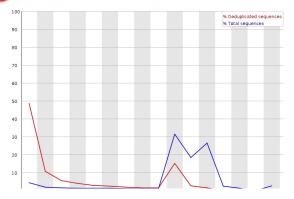
#### Basic Statistics

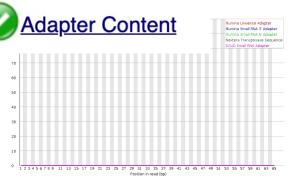
Measure	Value
Filename	forward.gz
File type	Conventional base calls
Encoding	Sanger / Illumina 1.9
Total Sequences	1264290
Sequences flagged as poor quality	0
Sequence length	76
%GC	39



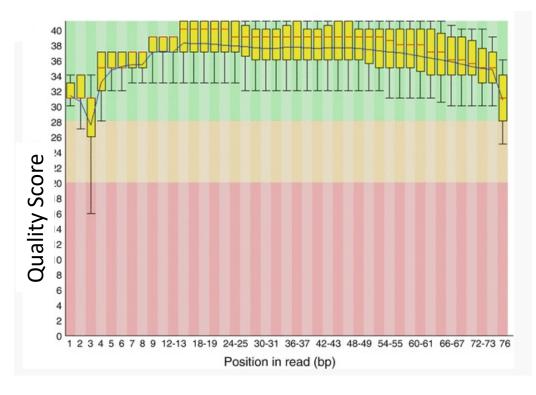


### Sequence Duplication Levels



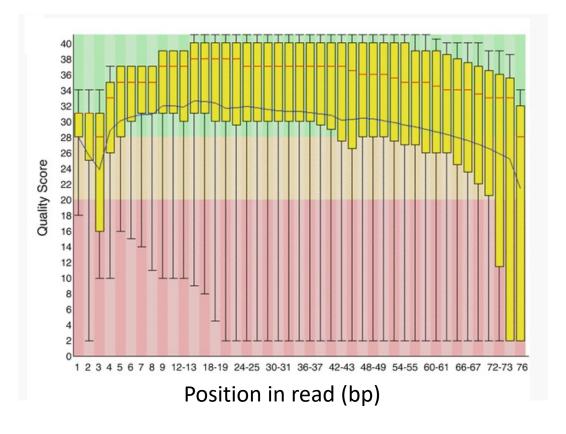


## FastQC: Sequence Quality Histogram

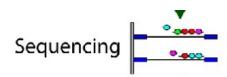


Position in read (bp)

GOOD
High quality over the length of the read

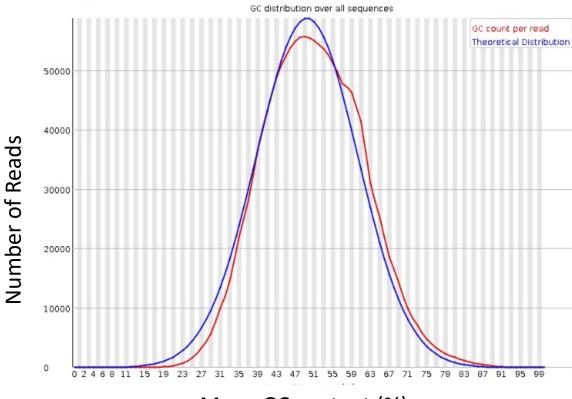


BAD
Read quality drops at the beginning and end

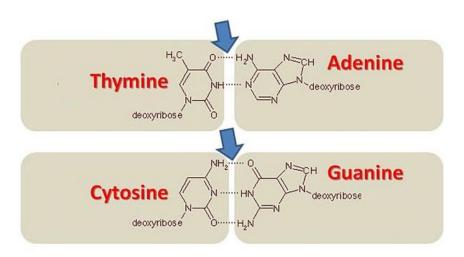


### FastQC: Per sequence GC content

#### Per sequence GC content

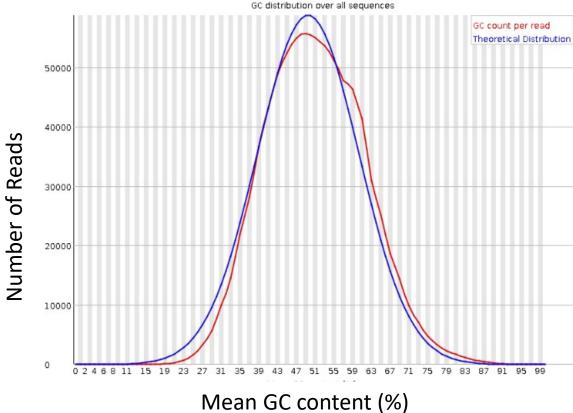


Mean GC content (%)



### FastQC: Per sequence GC content

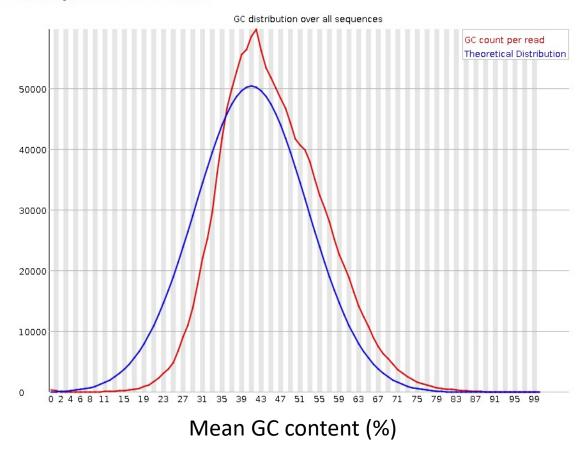
#### Per sequence GC content



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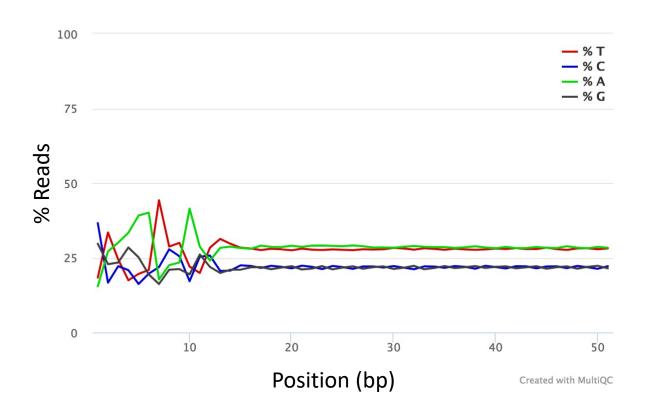
GOOD: follows normal distribution (sum of deviations is < 15% of reads)

#### **Per sequence GC content**



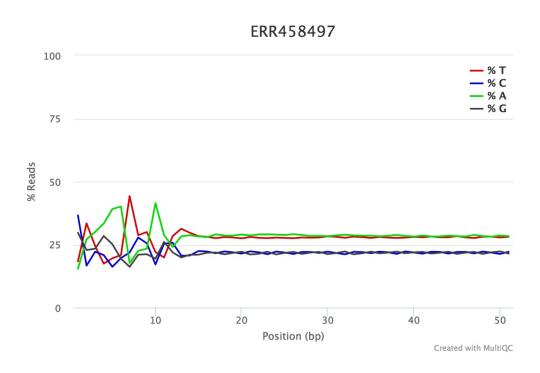
BAD: can indicate contamination with adapter dimers, or another species

## FastQC: Per Base Sequence Content

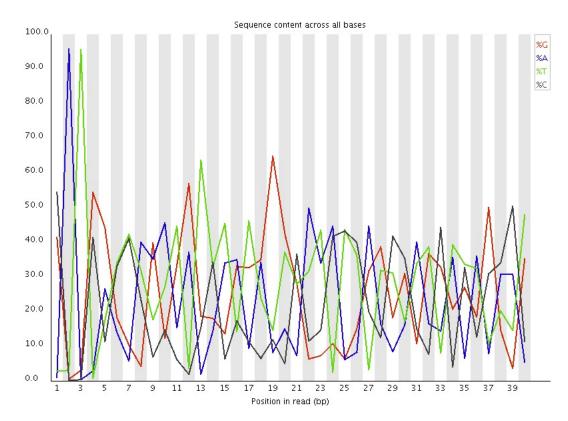


- Proportion of each position for which each DNA base has been called
- RNAseq data tends to show a positional sequence bias in the first ~12 bases
- The "random" priming step during library construction is not truly random and certain hexamers are more prevalent than others

### FastQC: Per Base Sequence Content



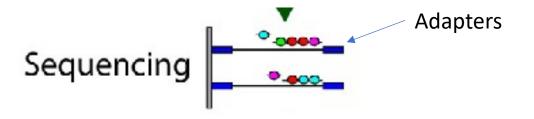
**EXPECTED** for RNAseq



BAD:

Shows a strong positional bias throughout the reads, which in this case is due to the library having a certain sequence that is overrepresented

## FastQC: Adapter content



FastQC will scan each read for the presence of known adapter sequences

The plot shows that the adapter content rises over the course of the read

Solution – Adapter trimming!

